# Partition Function and Base Pairing Probabilities for RNA-RNA Interaction Prediction 

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#### Abstract

The RNA-RNA interaction problems (RIP) deals with the energetically optimal structure of two RNA molecules that bind to each other. The standard model introduced by Alkan et al. (J. Comput. Biol. 13: 267-282, 2006) allows secondary structures in both partners as well as additional base pairs between the two RNAs subject to certain restrictions that allow a polynomial time dynamic programming solution. We derive the partition function for RIP based on a notion of "tight structures" as an alternative to the approach of Chitsaz et al. (Bioinformatics, 25, i365-i373, 2009). This dynamic programming approach is extended here by a fullfledged computation of the base pairing probabilities. The $O\left(N^{6}\right)$ time and $O\left(N^{4}\right)$ space algorithm is implemented in C (available from http://www.combinatorics.cn/cbpc/rip.html) and is efficient enough to investigate for instance the interactions of small bacterial RNAs and their target mRNAs.


## 1 INTRODUCTION

RNA-RNA interactions constitute one of the fundamental mechanisms of cellular regulation. In an important subclass, small RNAs specifically bind a larger (m)RNA target. Examples include the regulation of translation in both prokaryotes (Narberhaus and Vogel, 2007) and eukaryotes (McManus and Sharp, 2002; Banerjee and Slack, 2002), the targeting of chemical modifications (Bachellerie et al., 2002), and insertion editing (Benne, 1992), transcriptional control (Kugel and Goodrich, 2007). The common theme in many RNA classes, including miRNAs, siRNAs, snRNAs, gRNAs, and snoRNAs is the formation of RNA-RNA interaction structures that are more complex than simple sense-antisense interactions. The ability to predict the details of RNA-RNA interactions both in terms
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of the thermodynamics of binding in its structural consequences is a necessary prerequisite to understanding RNA based regulation mechanisms. The exact location of binding and the subsequent impact of the interaction on the structure of the target molecule can have profound biological consequences. In the case of sRNAmRNA interactions, these details decide whether the sRNA is a positive or negative regulator of transcription depending on whether binding exposes or covers the Shine-Dalgarno sequence (Sharma et al., 2007; Majdalani et al., 2002). Similar effects have been observed using artificially designed opener and closer RNAs that regulate the binding of the HuR protein to human mRNAs (Meisner et al., 2004; Hackermüller et al., 2005).

In its most general form, the RNA-RNA interaction problem (RIP) is NP-complete (Alkan et al., 2006; Mneimneh, 2007). The argument for this statement is based on an extension of the work of Akutsu (2000) for RNA folding with pseudoknots. Polynomialtime algorithms can be derived, however, by restricting the space of allowed configurations in ways that are similar to pseudoknot folding algorithms (Rivas and Eddy, 1999). The second major problem concerns the energy parameters since the standard loop types (hairpins, internal and multiloops) are insufficient; for the additional types, such as kissing hairpins, experimental data are virtually absent. Tertiary interactions, furthermore, are likely to have a significant impact.

Several restricted versions of RNA-RNA interaction have been considered in the literature. The simplest approach concatenates the two interacting sequences, essentially employing a slightly modified secondary structure folding algorithm. The algorithms RNAcofold (Hofacker et al., 1994; Bernhart et al., 2006), pairfold (Andronescu et al., 2005), and NUPACK (Ren et al., 2005) belong to this class. One major shortcoming of this approach is that it cannot predict important motifs such as kissing-hairpin loops. The paradigm of concatenation has also been generalized to the pseudoknot folding algorithm of Rivas and Eddy (1999).

The resulting model, however, still does not generate all relevant interaction structures (Chitsaz et al., 2009; Qin and Reidys, 2008). An alternative approach is to neglect all internal base pairings in either strand and to compute the minimum free energy (mfe) secondary structure for their hybridization under this constraint. For instance, RNAduplex and RNAhybrid (Rehmsmeier et al., 2004) follow this paradigm. RNAup (Mückstein et al., 2006, 2008) and intaRNA (Busch et al., 2008) restrict interactions to a single interval that remains unpaired in the secondary structure for each partner. These models have proved particularly useful for bacterial sRNA-mRNA interactions. Due to the highly conserved interaction motif, snoRNA-target interaction structures can be dealt with efficiently using specialized tools (Tafer et al., 2009).

Pervouchine (2004) and Alkan et al. (2006) independently derived and implemented mfe folding algorithms for predicting the joint secondary structure of two interacting RNA molecules with polynomial time complexity. In their model, a "joint structure" means that the intramolecular structures of each molecule are pseudoknot-free, the intermolecular binding pairs are noncrossing and there exist no so-called "zigzags", see Fig. 1(A) and 2(A) for examples of the "joint structures". The optimal "joint structure" can be computed in $O\left(N^{6}\right)$ time and $O\left(N^{4}\right)$ space by means of dynamic programming.

Recently, Chitsaz et al. (2009) presented piRNA, a tool that uses dynamic programming algorithm to compute the partition function of "joint structures", also in $O\left(N^{6}\right)$ time. The algorithmic cores of the forward recursions of piRNA and rip were developed independently. Albeit differing in design details, they are equivalent. In addition, we identified here a basic data structure that forms the basis for computing additional important quantities such as the base pairing probability matrix, and probabities of hybrid formations (see (Huang et al., 2009) for the latter). Further differences between the two approaches will be discussed in Section 5.

The key innovation for passing from the mfe folding of Alkan et al. (2006) to the partition function is a unique grammar by which each interaction structure can be generated. Then, the computation of the partition function follows McCaskill's approach for RNA secondary structures (McCaskill, 1990). The key idea is to identify a certain subclass of interaction structures that serve as building blocks in a recursive decomposition generalizing the loop decomposition of secondary structures. These are the "tight structures", a generalization of the subsecondary structures enclosed by a unique closing pair.

In the following two sections we first derive a grammar that allows the unambiguous parsing of zigzag-free interaction structures, thus forming the basis for the computation of the partition function in $O\left(N^{6}\right)$ time and $O\left(N^{4}\right)$ memory, corresponding the mfe algorithm of Alkan et al. (2006). Then we proceed by deriving the recursions for the base pairing probabilities, which are based on a conceptual reversing of the production rules. Indeed, one has to compute the pairing probabilities by explicitly "tracing back" all contributing joint structures. The output of rip consists of the partition function, the base pairing probability matrix and the joint structure predicted by the maximal weighted (in terms of the base pair probabilities) matching (MWM) algorithm (Cary and Stormo, 1995; Gabow, 1973) and the most likely hybrid loops.

The $\operatorname{sod} B-R h y B$ interaction structure (Geissmann and Touati, 2004) is a well-known paradigmatic example with a unique interaction region, Fig. 1. The rip software predicts this interaction
region correctly. Results obtained with other algorithms deviate noticeably from the known structure, see Supplemental Fig. S2.


Fig. 1. (A) The natural structure of $\operatorname{sodB}-R h y B$ (Geissmann and Touati, 2004). (B) The base pairing probability matrix (McCaskill, 1990) generated via rip. This matrix represents all potential base pairs of the $\operatorname{sod} B-R h y B$ structure as squares, whose area is proportional to their respective probability. Intermolecular and intramolecular base pairs are depicted in the blue upper right rectangle and the two white triangles, respectively. (C) "Zoom" into the most likely interaction region as predicted by rip. All base pairs of the hybrid are labeled by their probabilities.

To-date, only a handful of interaction structures are known that are more complex than those covered by intaRNA/RNAup. The best-known example is the repression of fhlA by OxyS RNA, which involves two widely separated kissing-hairpin loops (Argaman and Altuvia, 2000). In Fig. 2, we display the natural interaction structure as well as the output of rip, which predicts two distinct interaction regions. The left (red) one coincides exactly with the published structure, while the right (blue) one differs by only two base pairs. We shall return to the fhlA-OxyS prediction in more detail in the Discussion section.

## 2 JOINT STRUCTURES

Given two RNA sequences $R$ and $S$ (e.g. an antisense RNA and its target) with $N$ and $M$ vertices, we index the vertices such that $R_{1}$ is the $5^{\prime}$ end of $R$ and $S_{1}$ denotes the $3^{\prime}$ end of $S$. The edges of $R$ and $S$ represent the intramolecular base pairs. A pre-structure, $G(R, S, I)$, is a graph with the following properties:

1. $R, S$ are secondary structures (each nucleotide being paired with at most one other nucleotide via hydrogen bonds, without internal pseudoknots);
2. $I$ is a set of arcs of the form $R_{i} S_{j}$ without pseudoknots, i.e., if $R_{i_{1}} S_{j_{1}}$, $R_{i_{2}} S_{j_{2}} \in I$ where $i_{1}<i_{2}$, then $j_{1}<j_{2}$ holds.


Fig. 2. (A) The natural structure of $f$ hlA-OxyS (Chitsaz et al., 2009). (B) The base pairing probability matrix see the caption of Fig. 1 for notation. (C) "Zoom" into the two distinct, most likely interaction regions, as predicted by rip.

An arc is called exterior if it is of the form $R_{i} S_{j}$ and interior, otherwise. Let $G$ be a graph and $V$ be a subset of $G$-vertices. The (induced) subgraph of $G$ induced by $V$ has vertex set $V$ and contains all $G$-edges having both incident vertices in $V$. In particular, we use $S[i, j]$ to denote the subgraph of the pre-structure $G(R, S, I)$ induced by $\left\{S_{i}, S_{i+1}, \ldots, S_{j}\right\}$, where $S[i, i]=S_{i}$ and $S[i, i-1]=\varnothing$. In absence of interactions a pre-structure is a pair of induced secondary structures on $R$ and $S$, which we will refer to as a pair of segments. A segment $S\left[i_{1}, j_{1}\right]$ is called maximal if there is no segment, $S[i, j]$ strictly containing $S\left[i_{1}, j_{1}\right]$.

An interior arc $R_{i_{1}} R_{j_{1}}$ is an $R$-ancestor of the exterior $\operatorname{arc} R_{i} S_{j}$ if $i_{1}<$ $i<j_{1}$. Analogously, $S_{i_{2}} S_{j_{2}}$ is an $S$-ancestor of $R_{i} S_{j}$ if $i_{2}<j<j_{2}$. The sets of $R$-ancestors and $S$-ancestors of $R_{i} S_{j}$ are denoted by $A_{R}\left(R_{i} S_{j}\right)$ and $A_{S}\left(R_{i} S_{j}\right)$, respectively. We will also refer to $R_{i} S_{j}$ as a descendant of $R_{i_{1}} R_{j_{1}}$ and $S_{i_{2}} S_{j_{2}}$ in this situation. The $R$ - and $S$-ancestors of $R_{i} S_{j}$ with minimum arc-length are referred to as $R$ - and $S$-parents, see Fig. 3, (A). Finally, we call $R_{i_{1}} R_{j_{1}}$ and $S_{i_{2}} S_{j_{2}}$ dependent if they have a common descendant and independent, otherwise.

(B)


Fig. 3. (A) Ancestors and parents: for the exterior arc $R_{3} S_{4}$, we have the following ancestor sets $A_{R}\left(R_{3} S_{4}\right)=\left\{R_{1} R_{6}, R_{2} R_{4}\right\}$ and $A_{S}\left(R_{3} S_{4}\right)=$ $\left\{S_{2} S_{6}, S_{3} S_{5}\right\}$. In particular, $R_{2} R_{4}$ and $S_{3} S_{5}$ are the $R$-parent and $S$ parent respectively. (B) Subsumed and equivalent arcs: $R_{1} R_{8}$ subsumes $S_{1} S_{4}$ and $S_{5} S_{8}$. Furthermore, $R_{2} R_{5}$ is equivalent to $S_{1} S_{4}$.

Suppose there is an exterior arc $R_{a} S_{b}$ with ancestors $R_{i} R_{j}$ and $S_{i^{\prime}} S_{j^{\prime}}$. Then $R_{i} R_{j}$ is subsumed in $S_{i^{\prime}} S_{j^{\prime}}$, if for any $R_{k} S_{k^{\prime}} \in I^{\prime}, i<k<j$ implies $i^{\prime}<k^{\prime}<j^{\prime}$, see Fig. 3, (B). If $R_{i_{1}} R_{j_{1}}$ is subsumed in $S_{i_{2}} S_{j_{2}}$ and
vice versa, we call these arcs equivalent. A zigzag, is a subgraph containing two dependent interior $\operatorname{arcs} R_{i_{1}} R_{j_{1}}$ and $S_{i_{2}} S_{j_{2}}$ neither one subsuming the other, see Fig. 4, (A).
(A)

(B) 1


Fig. 4. (A): A zigzag, generated by $R_{2} S_{1}, R_{3} S_{3}$ and $R_{5} S_{4}$. (B): the joint structure $J_{1,24 ; 1,23}$, we display the different segments and tight structures in which $J_{1,24 ; 1,23}$ decomposes.

A joint structure, $J(R, S, I)$, is a zigzag-free pre-structure, see Fig. 4, (B). Joint structures are exactly the configurations that are considered in the maximum matching approach of Pervouchine (2004), in the energy minimization algorithm of Alkan et al. (2006), and in the partition function approach of Chitsaz et al. (2009). The subgraph of a joint structure $J(R, S, I)$ induced by a pair of subsequences $\left\{R_{i}, R_{i+1}, \ldots, R_{j}\right\}$ and $\left\{S_{h}, S_{h+1}, \ldots, S_{\ell}\right\}$ is denoted by $J_{i, j ; h, \ell}$. In particular, $J(R, S, I)=$ $J_{1, N ; 1, M}$. We say $R_{a} R_{b}\left(S_{a} S_{b}, R_{a} S_{b}\right) \in J_{i, j ; h, \ell}$ if and only if $R_{a} R_{b}\left(S_{a} S_{b}, R_{a} S_{b}\right)$ is an edge of the graph $J_{i, j ; h, \ell}$. Furthermore, $J_{i, j ; h, \ell} \subset J_{a, b ; c, d}$ if and only if $J_{i, j ; h, \ell}$ is a subgraph of $J_{a, b ; c, d}$ induced by $\left\{R_{i}, \ldots, R_{j}\right\}$ and $\left\{S_{h}, \ldots, S_{\ell}\right\}$.

We next define a tight structure (ts). Given a joint structure, $J_{a, b ; c, d}$, its tight $J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}$ is either a single exterior arc $R_{a^{\prime}} S_{c^{\prime}}$ (in the case $a^{\prime}=$ $b^{\prime}$ and $c^{\prime}=d^{\prime}$ ), or the minimal block centered around the leftmost and rightmost exterior arcs $\alpha_{l}, \alpha_{r}$, (possibly being equal) and an interior arc subsuming both, i.e., $J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}$ is tight in $J_{a, b ; c, d}$ if it has either an arc $R_{a^{\prime}} R_{b^{\prime}}$ or $S_{c^{\prime}} S_{d^{\prime}}$ if $a^{\prime} \neq b^{\prime}$ or $c^{\prime} \neq d^{\prime}$.
More formally, let $J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}$ be contained in $J_{a, b ; c, d}$ with rightmost and leftmost exterior $\operatorname{arc} R_{i} S_{j}$ and $R_{i_{0}} S_{j_{0}}$ and let $M$ be the set of $R_{i} S_{j}$ ancestors in $J_{a, b ; c, d}$ with maximal length. Then $J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}$ is tight in $J_{a, b ; c, d}$ if

1. for $M=\varnothing: J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}=\left\{R_{i} S_{j}\right\}$;
2. for $M=\left\{R_{i_{1}} R_{j_{1}}\right\}: J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}=J_{i_{1}, j_{1} ; c^{\prime}, j}$, where $c^{\prime}$ is the origin (left) of the $S$-ancestor of $R_{i_{0}}, S_{j_{0}}$ with maximal length (or $i_{0}$ if there is none). The case $M=\left\{S_{r_{1}} S_{s_{1}}\right\}$ is analogous;
3. for $M=\left\{R_{i_{1}} R_{j_{1}}, S_{r_{1}} S_{s_{1}}\right\}$, suppose $R_{i_{1}} R_{j_{1}}$ subsumes $S_{r_{1}} S_{s_{1}}$. Then $J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}=J_{i_{1}, j_{1} ; x_{1}, s_{1}}$, where $x_{1}$ is the origin of the $S$-ancestor of $R_{i_{0}} S_{j_{0}}$ with maximal length (or $i_{0}$ if there is none). In particular, $J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}=J_{i_{1}, j_{1} ; r_{1}, s_{1}}$ when $R_{i_{1}} R_{j_{1}}$ is equivalent with $S_{r_{1}} S_{s_{1}}$. The case, where $S_{r_{1}} S_{s_{1}}$ subsumes $R_{i_{1}} R_{j_{1}}$ is analogous.

In the following, a ts is denoted by $J_{i, j ; h, \ell}^{T}$. If $J_{a^{\prime}, b^{\prime} ; c^{\prime}, d^{\prime}}$ is tight in $J_{a, b ; c, d}$, then we call $J_{a, b ; c, d}$ its envelope. By construction, the notion of ts is depending on its envelope. There are only four basic types of ts, see Fig. 5:

$$
\begin{aligned}
& \circ:\left\{R_{i} S_{h}\right\}=J_{i, j ; h, \ell}^{\circ} \text { and } i=j, h=\ell ; \\
& \nabla: R_{i} R_{j} \in J_{i, j ; h, \ell}^{\nabla} \text { and } S_{h} S_{\ell} \notin J_{i, j ; h, \ell}^{\nabla} ; \\
& \square:\left\{R_{i} R_{j}, S_{h} S_{\ell}\right\} \in J_{i, j ; h, \ell}^{\square} ; \\
& \triangle: S_{h} S_{\ell} \in J_{i, j ; h, \ell}^{\triangle} \text { and } R_{i} R_{j} \notin J_{i, j ; h, \ell}^{\triangle} .
\end{aligned}
$$

In the Supplemental Material we prove:
Proposition 2.1. Let $J_{a, b ; c, d}$ be a joint structure. Then


Fig. 5. From left to right: tights of type $\circ, \nabla, \square$ and $\triangle$.

1. any exterior arc $R_{i} S_{j}$ in $J_{a, b ; c, d}$ is contained in a unique $J_{a, b ; c, d}-t s$;
2. $J_{a, b ; c, d}$ decomposes into a unique collection of $J_{a, b ; c, d^{-t s}}$ and maximal segments.

Given a ts, $J_{i_{0}, j_{0} ; r, s}^{\nabla}$ (or $J_{i, j ; r_{0}, s_{0}}^{\triangle}$ ), we introduce double tight structures as maximal substructure whose distinct leftmost and rightmost blocks are tights. By construction, therefore, each dts contains at least two tights.

More formally, given a ts $J_{i_{0}, j_{0} ; r, s}^{\nabla}$, a double-tight structure, $J_{i, j ; r, s}^{D T \mid \nabla}$, in $J_{i_{0}, j_{0} ; r, s}^{\nabla}$, where $i_{0}<i<j<j_{0}$, is defined as follows: there exists labels $a, b, c, d$ where $i \leq a<b \leq j$ and $r \leq c<d \leq s$. Furthermore, two ts $J_{i, a ; r, c}^{T}$ and $J_{b, j ; d, s}^{T}$ in $J_{i_{0}+1, j_{0}-1 ; r, s}$ such that

$$
\begin{equation*}
J_{i, j ; r, s}^{D T \mid \nabla}=J_{i, a ; r, c}^{T} \dot{\cup} J_{a+1, b-1 ; c+1, d-1} \dot{\cup} J_{b, j ; d, s}^{T} \tag{2.1}
\end{equation*}
$$

Here, the disjoint union $\dot{U}$ refers to both the vertex and arc sets of the joint structures, see Fig. 6. The case of a dts, $J_{i, j ; r, s}^{D T \mid \triangle}$, within a ts, $J_{i, j ; r_{0}, s_{0}}^{\triangle}$, is defined accordingly. By abuse of terminology, we simply use $J_{i, j ; r, s}^{D T}$ in order to denote either $J_{i, j ; r, s}^{D T \mid \nabla}$ or $J_{i, j ; r, s}^{D T \mid \triangle}$.


Fig. 6. A dts $J_{6,15 ; 1,11}^{D T \mid \nabla}$ in $J_{2,20 ; 1,11}$. Note that the joint structure $J_{1,21 ; 1,11}$ itself is $\nabla$-tight. Here, $J_{2,15 ; 1,11}$ is neither a ts nor a rts in $J_{2,20 ; 1,11}$.

With the help of dts, as illustrated in Fig. 7, Procedure (b), we decompose a ts as follows:
Let $J_{i, j ; r, s}^{\nabla}$ be a ts of type $\nabla$ and let $R_{h_{1}} S_{\ell_{1}}$ and $R_{h_{2}} S_{\ell_{2}}$ be the leftmost and rightmost exterior arcs in $J_{i, j ; r, s}$ and $i+1 \leq i_{1} \leq j_{1} \leq j-1$. Then $J_{i+1, j-1 ; r, s}$ decomposes into

$$
\left\{\begin{align*}
& R\left[i+1, i_{1}-1\right] \dot{\cup} J_{i_{1}, j_{1} ; r, s}^{\{\nabla, \circ\}} \dot{\cup} R\left[j_{1}+1, j-1\right]  \tag{2.2}\\
& \text { if } J_{R_{h_{1}} S_{\ell_{1}}}^{T}=J_{R_{h_{2}} S_{\ell_{2}}}^{T} \\
& R\left[i+1, i_{1}-1\right] \dot{\cup} J_{i_{1}, j_{1} ; r, s}^{D T} \dot{\cup} R\left[j_{1}+1, j-1\right] \\
& \text { otherwise, }
\end{align*}\right.
$$

where $J_{i_{1}, j_{1} ; r, s}^{\{\nabla, \circ\}}$ denotes a $J_{i+1, j-1 ; r, s}$-ts of type $\nabla$ or $\circ$ and $J_{R_{h}}^{T} S_{\ell}$ denotes the unique ts in $J_{i+1, j-1 ; r, s}$ contain the exterior $\operatorname{arc} R_{h} S_{\ell}$.

Analogously, in case of a ts $J_{i, j ; r, s}^{\triangle}$ with leftmost and rightmost exterior $\operatorname{arcs} R_{h_{1}} S_{\ell_{1}}$ and $R_{h_{2}} S_{\ell_{2}}$, and $r+1 \leq r_{1} \leq s_{1} \leq s-1, J_{i, j ; r+1, s-1}$ can be decomposed in the form

$$
\left\{\begin{array}{c}
S\left[r+1, r_{1}-1\right] \dot{\cup} J_{i, j ; r_{1}, s_{1}}^{\{\triangle, \circ\}} \dot{\cup} S\left[s_{1}+1, s-1\right]  \tag{2.3}\\
\text { if } J_{R_{h_{1}}, S_{\ell_{1}}}^{T}=J_{R_{h_{2}} S_{\ell_{2}}}^{T} \\
S\left[r+1, r_{1}-1\right] \dot{\cup} J_{i, j ; r_{1}, s_{1}}^{D T} \dot{\cup} S\left[s_{1}+1, s-1\right] \\
\text { otherwise, }
\end{array}\right.
$$

Kissing loops have been singled out for logical reasons and because some investigations into their thermodynamic properties have been reported in the literature (Gago et al., 2005). For details of the parametrization employed in rip we refer to the SM.

Let us now have a closer look at the energy evaluation of $J_{i, j ; h, \ell}$. Each decomposition step in Fig. 7 results in substructures whose energies we assume to contribute additively and generalized loops that need to be evaluated directly. There are the following two scenarios:

Procedure (a)


Procedure (b)


Fig. 7. Illustration of Procedure (a) the reduction of arbitrary joint structures and rts, and Procedure (b) the decomposition of tight structures. The panel below indicates the 10 different types of structural components: $\mathbf{A}, \mathbf{B}$ : maximal secondary structure segments $R[i, j], S[r, s]$; C: arbitrary joint structure $J_{i, j ; r, s} ; \mathbf{D}$ : right-tight structures $J_{i, j ; r, s}^{R T} ; \mathbf{E}$ : double-tight structure $J_{i, j ; r, s}^{D T} ; \mathbf{F}$ : tight structure of type $\nabla, \triangle$ or $\square$; $\mathbf{G}$ : type $\square$ tight $J_{i, j ; r, s}^{\square}$, the solid curved line (top and bottom) denotes an arc and a single horizontal line (top and bottom) denotes the backbone; $\mathbf{H}$ : type $\nabla$ tight $J_{i, j ; r, s}^{\nabla}$, a solid curved line (top) denotes an arc, a single horizontal line (top) denotes the backbone and a double-horizontal line (bottom) denotes that the two terminals are not paired wit each other; $\mathbf{J}$ : type $\triangle$ tight $J_{i, j ; r, s}^{\triangle}$, a solid curved line (bottom) denotes an arc, a single horizontal line (bottom) denotes the backbone and a double-horizontal line (top) indicates that the two terminals are not paired with each other; $\mathbf{K}$ : an exterior arc.


Fig. 8. The decomposition tree $T_{J_{1,15 ; 1,8}}$ for the joint structure $J_{1,15 ; 1,8}$.


Fig. 9. The two new loop-types: hybrid (A) and kissing-loop (B).
I. Arc removal. Most of the decomposition operations in Procedure (b) displayed in Fig. 7 can be viewed as the "removal" of an arc (corresponding to the closing pair of a loop in secondary structure folding) followed by decomposition. Both, loop-type as well as the subsequent decomposition steps depend on the newly exposed structural elements. Following the approach of Zuker and Stiegler (1981) for secondary structures, we treat the
loop-decomposition problem by introducing additional matrices. Without loss of generality, we can assume that we open an interior base pair $R_{i} R_{j}$.

The set of base pairs on $R[i, j]$ consists of all interior pairs $R_{p} R_{q}$ with $i \leq p<q \leq j$ and all exterior pairs $R_{p} S_{h}$ with $i \leq p \leq j$. An interior arc is exposed on $R[i+1, j-1]$ if and only if it is not enclosed by any interior arc in $R[i, j]$. An exterior arc is exposed on $R[i+1, j-1]$ if and only if it is not a descendant of any interior arc in $R[i+1, j-1]$. Given $R_{i j}$, the arcs exposed on $R[i+1, j-1]$ corresponds to the base pairs immediately interior of $R_{i} R_{j}$. Let us write $E_{R[i, j]}=E_{R[i, j]}^{i} \dot{\cup} E_{R[i, j]}^{e}$ for this set of "exposed base pairs" and its subsets of interior and exterior arcs. As in secondary structure folding, the loop type is determined by $E_{R[i, j]}:=E_{R}$ as follows: $E_{R}=\varnothing$, hairpin loop; $E_{R}=E_{R}^{i}$ and $\left|E_{R}\right|=1$, interior loop (including bulge and stacks); $E_{R}=E_{R}^{i},\left|E_{R}\right| \geq 2$, multi-branch loop; $E_{R}=E_{R}^{e}$, kissing-hairpin loop; $\left|E_{R}^{i}\right|,\left|E_{R}^{e}\right| \geq 1$, general kissing-loop.
This picture needs to be refined even further since the arc removal is coupled with further decomposition of the interval $R[i+1, j-1]$. This prompts us to distinguish ts and dts with different classes of exposed base pairs on one or both strands. It will be convenient, furthermore to include information on the type of loop in which it was found.

A ts $J_{i, j ; h, \ell}^{\nabla}$ is of type E , if $S[h, \ell]$ is not enclosed in any base pair $\left(J_{i, j ; h, \ell}^{\nabla, \mathrm{E}}\right)$. Suppose $J_{i, j ; h, \ell}^{\nabla}$ is located immediately interior to the closing pair $S_{p} S_{q}(p<h<\ell<q)$. If the loop closed by $S_{p} S_{q}$ is a multiloop, then $J_{i, j ; h, \ell}^{\nabla}$ is of type $\mathrm{M}\left(J_{i, j ; h, \ell}^{\nabla, \mathrm{M}}\right)$. If $S_{p} S_{q}$ is contained in a kissing-loop, we distinguish the types F and K , depending on whether or not $E_{S[h, \ell]}^{e}=\varnothing$. Fig. 10 displays this decomposition for $J_{i, j ; r, s}^{\nabla, \mathrm{M}}$.


Fig. 10. Further refinement: the four decompositions of $J_{i, j ; r, s}^{\nabla, M}$ via Procedure (b). The green rectangle denotes single-stranded segments. The letters I, M, etc denote the loop-type and the type of the exposed arc(s) of the dts. See Fig. 7 for more details on the notation. The four cases correspond to the four contributions in equ. (3.1).

For a dts $J_{p, q, r, s}^{D T}$ (denoted by " E " in Fig. 7) we need to determine the type of the exposed pairs of both $R[p, q]$ and $S[r, s]$. Hence each such structure will be indexed by two types. In total, we arrive at 18 distinct cases since some combinations cannot occur. For instance, a dts cannot be external in both $R$ and $S$, that is, type EE does not exist, where E means external.


Fig. 11. Decomposition of $J_{i, j ; h, \ell}^{R T, K K B}$ by means of procedure (b). Here the red rectangle denotes a pair of secondary segments having the property that at least one of them is not single-stranded.


Fig. 12. Decomposition of $J_{i, j ; h, \ell}^{D T, \mathrm{KM}}$ by means of procedure (b). The five alternatives correspond to the three additive terms in equ. (3.2).
II. Block decomposition. The second type of decomposition is the splitting of joint structures into "blocks", such as the decompositions of a rts in Procedure (a) and a dts in Procedure (b), see Fig. 7. A rts $J_{i, j ; h, \ell}^{R T}$ may appear in two ways, depending on whether or not there exists an exterior arc $R_{i_{1}} S_{j_{1}}$ such that $R\left[i_{1}+1, j\right]$ and $S\left[j_{1}+1, \ell\right]$ are single-stranded segments. If such an exterior arc exists, $J_{i, j ; h, \ell}^{R T}$ is of type (rB), otherwise it is of type (rA). Analogously, $(\ell B)$ and ( $\ell A$ ) are defined for dts. Fig. 11 shows the decomposition of $J_{i, j ; h, \ell}^{R T, \mathrm{KKB}}$. Suppose $J_{i, j ; r, \ell}^{D T}$ is a dts contained in a kissingloop, that is we have either $E_{R[i, j]}^{e} \neq \varnothing$ or $E_{S[h, \ell]}^{e} \neq \varnothing$. W.l.o.g., we may assume $E_{R[i, j]}^{e} \neq \varnothing$. Then at least one of the two "blocks" contains the exterior arc belonging to $E_{R[i, j]}^{e}$ (i.e., direct bonds in the language of Chitsaz et al. (2009)) labeled by K. Otherwise the block is labeled F, see Fig. 12. The situation is analogous if we decompose $J_{i, j ; r, \ell}^{R T}$ contained in a kissing-loop.

### 3.3 Recursions

The computation of the partition function is obtained "from the inside to the outside", see equs. (3.1,3.2). The recursions are initialized with the energies of individual external base pairs and empty secondary structures on subsequences of length up to four. In order to differentiate multi- and kissing-loop contributions, we introduce the partition functions $Q_{i, j}^{\mathrm{m}}$ and $Q_{i, j}^{\mathrm{F}}$ as a generalization of Zuker's algorithm (Zuker and Stiegler, 1981). Here, $Q_{i, j}^{\mathrm{m}}$ denotes the partition function of secondary structures on $R[i, j]$ or $S[i, j]$ having at least one arc contained in a multi-loop. Similarly, $Q_{i, j}^{\mathrm{F}}$ denotes the partition function of secondary structures on $R[i, j]$ or $S[i, j]$ in which at least one arc is contained in a kissing-loop.

For instance, the recursion for $Q_{i, j ; r, s}^{\nabla, \mathrm{M}}$ in Fig. 10 reads:

$$
\begin{align*}
Q_{i, j ; r, s}^{\nabla, \mathrm{M}}= & \sum_{h, \ell}\left\{Q_{h, \ell ; r, s}^{\nabla, \mathrm{M}} e^{-G_{i, j ; h, \ell}^{\mathrm{Int}} / k T}\right. \\
+ & Q_{h, \ell ; r, s}^{D T, \mathrm{MM}} e^{-\left(\alpha_{1}+\alpha_{2}\right) / k T} \times\left(e^{-(h-i-1) \alpha_{3} / k T}+Q_{i+1, h-1}^{\mathrm{m}}\right) \\
& \times\left(e^{-(j-\ell-1) \alpha_{3} / k T}+Q_{\ell+1, j-1}^{\mathrm{m}}\right) \\
+ & Q_{h, \ell ; r, s}^{D T, \mathrm{KM}} e^{-\left(\beta_{1}+\beta_{2}\right) / k T} \times\left(e^{-(h-i-1) \beta_{3} / k T}+Q_{i+1, h-1}^{\mathrm{F}}\right) \\
& \times\left(e^{-(j-\ell-1) \beta_{3} / k T}+Q_{\ell+1, j-1}^{\mathrm{F}}\right), \\
+ & Q_{h, \ell ; r, s}^{\nabla, \mathrm{M}} e^{-\left(\alpha_{1}+2 \alpha_{2}\right) / k T}\left[e^{-(j-\ell-1) \alpha_{3} / k T} Q_{i+1, h-1}^{\mathrm{m}}\right. \\
& \left.\left.+e^{-(h-i-1) \alpha_{3} / k T} Q_{\ell+1, j-1}^{\mathrm{m}}+Q_{\ell+1, j-1}^{\mathrm{m}} Q_{i+1, h-1}^{\mathrm{m}}\right]\right\} \tag{3.1}
\end{align*}
$$

Analogously, the recursion for the dts $Q_{i, j ; r, s}^{D T, \mathrm{KM}}$ of Fig. 12 is given by

$$
\begin{align*}
Q_{i, j ; r, s}^{D T, \mathrm{KM}} & =\sum_{i_{1}, j_{1}}\left\{\left(Q_{i, i_{1} ; r, j_{1}}^{\nabla, \mathrm{M}} e^{-\beta_{2} / k T}+Q_{i, i_{1} ; r, j_{1}}^{\triangle, \mathrm{K}} e^{-\alpha_{2} / k T}\right.\right. \\
& \left.+Q_{i, i_{1} ; r, j_{1}}^{\square} e^{-\left(\alpha_{2}+\beta_{2}\right) / k T}+Q_{i, i_{1} ; r, j_{1}}^{\triangle, \mathrm{F}} e^{-\alpha_{2} / k T}\right) Q_{i_{1}+1, j ; j_{1}+1, s}^{R T, \mathrm{KM}} \\
& \left.+Q_{i, i_{1} ; r, j_{1}}^{\triangle, \mathrm{K}} e^{-\alpha_{2} / k T} Q_{i_{1}+1, j ; j_{1}+1, s}^{R T T, \mathrm{FM}}\right\} . \tag{3.2}
\end{align*}
$$

## 4 BASE PAIRING PROBABILITIES

Given two RNA sequences, our sample space is the ensemble of all zigzagfree joint interaction structures. Let $Q^{I}$ denote the corresponding partition function. The probability of a joint structure $J_{1, N ; 1, M}$ is then given by $\mathbb{P}_{J_{1, N ; 1, M}}=Q_{J_{1, N ; 1, M}} / Q^{I}$.

### 4.1 Approach

While the computation of the partition function proceeds from smaller to larger subsequences, the computation of the substructure probabilities follows the order of the decomposition outlined in the previous section. That is, the longest-range substructures are computed first, analogous to McCaskill's algorithm for secondary structures (McCaskill, 1990).

Let $\mathbb{J}_{i, j ; h, \ell}^{\xi, Y_{1} Y_{2} Y_{3}}$ be the set of substructures $J_{i, j ; h, \ell} \subset J_{1, N ; 1, M}$ such that $J_{i, j ; h, \ell}$ appears in $T_{J_{1, N ; 1, M}}$ as an interaction structure of type $\xi \in$ $\{D T, R T, \nabla, \triangle, \square, \circ\}$ with loop-subtypes $Y_{1}, Y_{2} \in\{\mathrm{M}, \mathrm{K}, \mathrm{F}\}$ on the sub-intervals $R[i, j]$ and $S[h, \ell], Y_{3} \in\{\mathrm{~A}, \mathrm{~B}\}$. Let $\mathbb{P}_{i, j ; h, \ell}^{\xi, Y_{1} Y_{2} Y_{3}}$ be the probability of $\mathbb{J}_{i, j ; h, \ell}^{\xi, Y_{1} Y_{2} Y_{3}}$. For instance, $\mathbb{P}_{i, j ; h, \ell}^{R T, \mathrm{MKA}}$ is the sum over all the probabilities of substructures $J_{i, j ; h, \ell} \in T_{J_{1, N ; 1, M}}$ such that $J_{i, j ; h, \ell}$ is a rts of type $r \mathrm{~A}$ and $R[i, j], S[h, \ell]$ are enclosed by a multi-loop and kissingloop, respectively. Then the computation of the pairing probabilities reduces to a trace-back routine in the decomposition tree constructed in Section 3.1.

Set $J=J_{1, N ; 1, M}, T=T_{J_{1, N ; 1, M}}$ and let $\Lambda_{J_{i, j ; h, \ell}}=\left\{J \mid J_{i, j ; h, \ell} \in\right.$ $T\}$ denote the set of all joint structures $J$ such that $J_{i, j ; h, \ell}$ is a vertex in the decomposition tree $T$. Then we have $\mathbb{P}_{J_{i, j ; h, \ell}}=\sum_{J \in \Lambda_{i, j ; h, \ell}} \mathbb{P}_{J}$ and furthermore

$$
\begin{equation*}
\mathbb{P}_{i, j ; h, \ell}^{\xi, Y_{1} Y_{2} Y_{3}}=\sum_{J_{i, j ; h, \ell} \in J_{i, j ; h, \ell}^{\xi, Y_{1} Y_{2} Y_{3}}} \mathbb{P}_{i, j ; h, \ell} . \tag{4.1}
\end{equation*}
$$

### 4.2 Case Study: Secondary Structures

In order to illustrate the logic of our backtracking procedure, we first consider the special case of secondary structures.
Let $\mathbb{P}_{R_{i} R_{j}}$ denote the base pairing binding probability of $R_{i} R_{j}$, i.e. $\mathbb{P}_{R_{i} R_{j}}=\sum_{R_{i} R_{j} \in W} Q_{W} Q^{-1}$, where the sums is taken over all the partition functions of secondary structures $W$ in $R$ such that $R_{i} R_{j} \in W$. Let $T_{W}$ be the decomposition tree of a particular secondary structure $W$ on $R[1, N]$ via Procedure (c), the key observation here is

$$
\begin{equation*}
R_{i} R_{j} \in W \Longleftrightarrow R_{i} R_{j} \in T_{W} \tag{4.2}
\end{equation*}
$$

Let $\Omega\left(R_{i} R_{j}\right)$ be the set of secondary structures whose decomposition tree contain the pair $R_{i} R_{j}$ as a leaf. Clearly, via equ. (4.2), we obtain

$$
\begin{equation*}
\mathbb{P}_{R_{i} R_{j}}=\sum_{W \in \Omega\left(R_{i} R_{j}\right)} Q_{W} Q^{-1} \tag{4.3}
\end{equation*}
$$

In order to compute $\mathbb{P}_{R_{i} R_{j}}$, we express it as a sum over the probabilities of all substructures $\xi$, that are a parent of $R_{i} R_{j}$ in the decomposition tree. Let $R^{b}(i, j)$ denote the set of secondary segments $R[i, j]$ in which $R_{i}$ is connected with $R_{j}$ and let $\mathbb{P}_{R_{i}, R_{j}}^{b}$ be its probability. By construction, we have $\mathbb{P}_{R_{i} R_{j}}=\mathbb{P}_{R_{i}, R_{j}}^{b}$, since the parent of $R_{i} R_{j}$ in the decomposition tree must be a secondary segment $R[i, j]$ such that $R_{i} R_{j} \in R[i, j]$. Therefore
the computation of $\mathbb{P}_{R_{i} R_{j}}$ is reduced to the calculation of the substructure probability $\mathbb{P}_{R_{i}, R_{j}}^{b}$.
The decomposition is summarized in Procedure (c), Fig. 13. In view of the fact that the rip has $O\left(N^{6}\right)$ time complexity, we can differ here from the standard implementation of the RNA folding model. Inspection of Fig. 13 shows that for a $R^{b}(i, j)$-parent we have to distinguish the five cases displayed in the lower panel. Let $R^{\mathrm{m}}(i, j)$ be the set of segments $R[i, j] \in T_{R[1, N]}$ containing at least one arc with an outer loop of type M , and let $R^{s}(i, j)$ be the set of all segments $R[i, j] \in T_{R[1, N]}$. Furthermore, let $\mathbb{P}_{R_{i}, R_{j}}^{\mathrm{m}}$ and $\mathbb{P}_{R_{i}, R_{j}}^{s}$ be the corresponding probabilities. Note that it is possible, for (L1) and (L4) in Fig. 13, that $h=i$ and $j=\ell$ holds. However, via further backtracking for $R^{s}(i, j)$ and $R^{m}(i, j)$ we can recursively calculate the binding probability.

Following the logic of Fig. 13, we obtain

$$
\begin{align*}
& \mathbb{P}_{R_{i}, R_{j}}^{b}=\sum_{h, \ell}\left\{\mathbb{P}_{R_{h}, R_{\ell}}^{s} \frac{Q_{h, i-1}^{s} Q_{i, j}^{b}}{Q_{h, \ell}^{s}}+\mathbb{P}_{R_{h}, R_{\ell}}^{b} \frac{Q_{i, j}^{b} e^{-G_{h, \ell ; i, j}^{\text {Int } / k T}}}{Q_{h, \ell}^{b}}\right. \\
& +\mathbb{P}_{R_{h}, R_{\ell}}^{b} \frac{Q_{h+1, i-1}^{\mathrm{m}} Q_{i, j}^{b} e^{-\left(\alpha_{1}+2 \alpha_{2}+(\ell-j-1) \alpha_{3}\right) / k T}}{Q_{h, \ell}^{b}} \\
& +\mathbb{P}_{R_{h}, R_{\ell}}^{\mathrm{m}} \frac{Q_{i, j}^{b} e^{-\left(\alpha_{2}+(i-h+\ell-j) \alpha_{3}\right) / k T}}{Q_{h, \ell}^{\mathrm{m}}} \\
& \left.+\mathbb{P}_{R_{h}, R_{\ell}}^{\mathrm{m}} \frac{Q_{h, i-1}^{\mathrm{m}} Q_{i, j}^{b} e^{-\left(\alpha_{2}+(\ell-j) \alpha_{3}\right) / k T}}{Q_{h, \ell}^{\mathrm{m}}}\right\} . \tag{4.4}
\end{align*}
$$

where the lines correspond to the five loop types (L1-L5) in Fig. 13. Analogously, the recursions for the base pairing probabilities $\mathbb{P}_{R_{i}, R_{j}}^{m}$ and $\mathbb{P}_{R_{i}, R_{j}}^{s}$ are given by

$$
\begin{align*}
& \mathbb{P}_{R_{i}, R_{j}}^{\mathrm{m}}=\sum_{h, \ell}\left\{\mathbb{P}_{R_{i-1}, R_{\ell}}^{b} e^{-\left(\alpha_{1}+2 \alpha_{2}+(\ell-1-h) \alpha_{3}\right) / k T}\right. \\
& \left.\quad \times \frac{Q_{j+1, h}^{b} Q_{i, j}^{\mathrm{m}}}{Q_{i-1, \ell}^{b}}+\mathbb{P}_{R_{i}, R_{\ell}}^{\mathrm{m}} \frac{\left.Q_{i, j}^{\mathrm{m}} Q_{j+1, h}^{b} e^{-\left(\alpha_{2}+(\ell-h) \alpha_{3}\right.}\right) / k T}{Q_{i, \ell}^{\mathrm{m}}}\right\} \\
& \mathbb{P}_{R_{i}, R_{j}}^{s}=\sum_{h, \ell} \mathbb{P}_{R_{i}, R_{\ell}}^{s} \frac{Q_{i, j}^{s} Q_{j+1, h}^{b}}{Q_{i, \ell}^{s}} \tag{4.5}
\end{align*}
$$

### 4.3 Base pairing probabilities for joint structures

Set $\Sigma_{1}=\left\{J \mid R_{i} R_{j} \in J\right\}$. We apply the same strategy to the joint structures appearing in Fig. 7. Let $Q^{I}$ denote the partition function which sums over all the possible joint structures $J_{1, N ; 1, M}$. Then $\mathbb{P}_{R_{i}, R_{j}}=$ $\sum_{J \in \Sigma_{1}} Q_{J} / Q^{I}$. In order to compute $\mathbb{P}_{R_{i} R_{j}}$ we classify $\Sigma_{1}$ according to the parent of $R_{i} R_{j}$ in $T$ :

$$
\begin{align*}
\Sigma_{1} & =\left\{J \mid R[i, j] \in T, R[i, j] \in R^{b}(i, j)\right\} \\
& \cup \bigcup_{h, \ell}\left\{J \mid J_{i, j ; h, \ell} \in T, J_{i, j ; h, \ell} \in \mathbb{J}_{i, j ; h, \ell}^{\nabla}\right\}  \tag{4.6}\\
& \cup \bigcup_{h, \ell}\left\{J \mid J_{i, j ; h, \ell} \in T, J_{i, j ; h, \ell} \in \mathbb{J}_{i, j ; h, \ell}^{\square}\right\}
\end{align*}
$$

which translates to

$$
\begin{equation*}
\mathbb{P}_{R_{i} R_{j}}=\mathbb{P}_{R_{i}, R_{j}}^{b}+\sum_{h, \ell} \mathbb{P}_{i, j ; h, \ell}^{\nabla,\{\mathrm{E}, \mathrm{M}, \mathrm{~F}, \mathrm{~K}\}}+\sum_{h, \ell} \mathbb{P}_{i, j ; h, \ell}^{\square}, \tag{4.7}
\end{equation*}
$$



Fig. 13. Top. Extended version of Procedure (c) showing the productions for general structures, structures and enclosed by pairs, and multi-loops. The four types of segments are shown below: In $R^{b}(i, j), R_{i}, R_{j}$ is paired, $R^{u}(i, j)$ denotes stretches of unpaired bases, $R^{\mathrm{m}}(i, j)$ denotes parts of multiloops with containing least one arc, and $R^{s}(i, j)$ denotes arbitrary segments. Below. Backtracking for secondary structures: for a parent of $R^{b}(i, j)$ we have five cases according to Procedure (c): external (L1), interior loop (L2), closing pair of a multi-loop (L3), (L4) and (L5) denote the scenarios arising from decomposing a $R^{m}(h, \ell)$-segment. See equ. (4.4) for the corresponding recursions.
where $\mathbb{P}_{i, j ; h, \ell}^{\nabla,\{\mathrm{E}, \mathrm{M}, \mathrm{F}, \mathrm{K}\}}=\mathbb{P} \nabla, \mathrm{E}+\mathbb{P} \nabla, \mathrm{M}+\mathbb{P} \nabla, \mathrm{F}+\mathbb{P} \nabla, \mathrm{K}$ for identical positions $i, j, h, \ell$. Analogously, we obtain for pairs in $S$ :

$$
\begin{align*}
& \Sigma_{2}=\left\{J \mid S[h, \ell] \in T, S[h, \ell] \in S^{b}[h, \ell]\right\} \\
& \cup \bigcup_{i, j}\left\{J \mid J_{i, j ; h, \ell} \in T, J_{i, j ; h, \ell} \in \mathbb{J}_{i, j ; h, \ell}^{\triangle}\right\} \tag{4.8}
\end{align*}
$$

and therefore $\mathbb{P}_{S_{i} S_{j}}=\mathbb{P}_{S_{i}, S_{j}}^{b}+\sum_{h, \ell} \mathbb{P}_{h, \ell ; i, j}^{\triangle}$, with $\mathbb{P}^{\triangle}=\mathbb{P}^{\triangle, \mathrm{E}}+$ $\mathbb{P}^{\triangle, M}+\mathbb{P}^{\triangle, K}+\mathbb{P}^{\triangle, F}$.

Note that the expressions for $\mathbb{P}_{R_{i} R_{j}}$ and $\mathbb{P}_{S_{i} S_{j}}$ are not symmetric. This is due to the fact that our decomposition routine give preference to arcremovals in $R$ over those in $S$. This asymmetry is necessary to ensure that the decomposition in Fig. 7 is unambiguous.

Finally, we calculate the binding probability of an exterior arc $R_{i} S_{j}$. Since $R_{i} S_{j}$ is a ts of type $\circ, \mathbb{P}_{R_{i} S_{j}}$ is directly given by the probability of this special substructure in equ. (4.1).

In order to compute the binding probabilities of both interior and exterior arcs, the key is to employ an "inverse" grammar induced by tracing back in the decomposition tree as displayed in Supplement Materials [SM, Fig. 5].

## 5 RESULTS AND DISCUSSION

In this contribution we have introduced a framework in which both the partition function and the base pairing probabilities of zigzag-free RNA-RNA interactions can be derived in a natural
way. Our approach is implemented in the software package rip using the full standard energy model for RNA secondary structures together with a multi-loop-like additive parametrization for kissingloops. In comparision with piRNA (Chitsaz et al., 2009), rip is based on a different but equivalent decomposition grammar. The very encouraging data on the accuracy of the predicted interaction intergies reported by Chitsaz et al. (2009) therefore carry over to rip as well.
The notion of tights, which have a central role in our presentation, is also implicit in the work of Chitsaz et al. (2009). The focus on the underlying combinatorial aspects, however, leads us to highlight in particular the decomposition tree, which provides a natural framework in which to proceed beyond the algorithmic core of the partition function itself. Indeed, the decomposition tree facilitates the derivation of the base pairing probabilities. Related questions, such as that for the probability of complete hybrids (Huang et al., 2009) can be answered along the same lines. While the current implmenentation of piRNA Chitsaz et al. (2009) concentrates on melting temperature in order to validate the partition function, rip focusses on a detailed analysis of the interaction structures themselves. To this end, we also compute the unweighted maximum expected accuracy structure, which is given as the maximum matching with weights given by the base pairing probabilites. On a more technical level, piRNA and rip differ in the decomposition of tights: piRNA utilizes a $4 D$ gap-matrix by means of the DirksPierce algorithm Dirks and Pierce (2003), while rip employs two distinct $2 D$-matrices inspired by Zuker's recursion. For details, we refer to the SM.

Back-tracing of the base pairing patterns that underlie the free energy of RNA-RNA binding is of great importance in detailed studies of ncRNA-mRNA interactions. The details of the binding sites have a crucial impact on the interpretation of the computational results and on the comparison of the computational prediction and experimental data. It was shown by Mückstein et al. (2008), for instance, that positive and negative regulation of bacterial mRNAs can be distinguished depending on whether the interaction structure contains the Shine-Dalgarno sequence in stable stem or exposed in an predominantly unpaired region.

Only a small number of interaction structures have been described so far that are more complex than those computable by RNAup/intaRNA. It is not clear, however, whether complex interactions are truely rare in nature, or whether multi-point contacts such as that of the fhlA-OxyS interaction structure (Argaman and Altuvia, 2000) are rarely observed experimentally because they are typically excluded from candidate lists due to the lack of readily detectable pairing regions. A survey with rip may be suitable to provide us with a much more unbiased picture. Fig. 2 shows, that, modulo two base pairs, rip identifies the two distinct hybrids in fhlA-OxyS, correctly. Table 1 furthermore establishes, that the latter are indeed uniquely identified.

Fig. 14 compares the output of rip with several other, established, folding algorithms.

Following Chitsaz et al. (2009), we stipulate an independent initialization energy, $\sigma_{0}$, for each hybrid, and scaled energies for its base pairs. Many other RNA-cofolding algorithms, like RNAcofold (Bernhart et al., 2006) and Dimitrov and Zuker (2004) assume a single initialization energy, $\varepsilon$. This energy model can a posteriori be derived from rip, once the partition function of the joint structures $Q^{I}$ and the partition functions $Q_{R}$ and $Q_{S}$

| I | II | III | IV |
| :---: | :---: | :---: | :---: |
| $58,47: 50.3 \%$ | $21,69: 56.5 \%$ | $34,55: 45.7 \%$ | $83,45: 17.3 \%$ |
| $59,46: 54.8 \%$ | $20,70: 59.5 \%$ | $35,54: 51.0 \%$ | $82,46: 18.9 \%$ |
| $60,45: 52.9 \%$ | $19,71: 29.8 \%$ | $36,53: 49.9 \%$ | $81,47: 18.5 \%$ |
| $61,44: 28.8 \%$ |  |  |  |

Table 1. The base pairing probabilities of the four alternative hybrids I, II, III and IV for fhlA-OxyS, predicted by rip, see Fig. 2. Each entry represents the positions in $f h l A(R)-O x y S(S)$ and the base pair probability. For instance, $58,47: 50.3 \%$ is equivalent to $\mathbb{P}_{R_{58} S_{47}}=$ 0.503 .


Fig. 14. The natural structure of the $f h l A-O x y S$ interaction and the results predicted by several algorithms including RNAplex.
have been computed. Let $\Omega_{1}$ be the set of all joint structures having at least one external arc and denote by $\Omega_{0}$ the set of all structures that have none. Then $Q^{\text {rip }}=Q\left(\Omega_{1}\right)+Q\left(\Omega_{0}\right)$, where $Q\left(\Omega_{0}\right)=Q_{R} Q_{S}$. Taking the initiation term into account, we compute $Q=Q\left(\Omega_{1}\right) \exp (-\varepsilon / k T)+Q_{R} Q_{S}$, from which we obtain the corrected value for $Q\left(\Omega_{1}\right)$. As shown by Bernhart et al. (2006), the base pairing probabilities can be rescaled via

$$
\begin{equation*}
\mathbb{P}_{i j}^{\epsilon}=\frac{\left[\mathbb{P}_{i j}^{\text {rip }} Q^{\text {rip }}-\mathbb{P}_{i j}\left(\Omega_{0}\right) Q_{R} Q_{S}\right] e^{-\varepsilon / k T}+\mathbb{P}_{i j}\left(\Omega_{0}\right) Q_{R} Q_{S}}{\left[Q^{\text {rip }}-Q_{R} Q_{S}\right] e^{-\varepsilon / k T}+Q_{R} Q_{S}} \tag{5.1}
\end{equation*}
$$

We have focussed here on the algorithmic context for computing detailed models of RNA-RNA interactions in the most general framework that is computationally feasible at the moment. The current implementation of rip may, due to the computational costs incurred by several dozens of interdependent 4-dimensional arrays, be viewed "just" as a reference. However, in all computed examples, rip quite accurately reproduced the interaction regions. We are here in a similar position as with the Sankoff algorithm (which addresses the closely related dynamic programming problem of simultaneous alignment and structure prediction). While the full implementations are slow and of limited use in particular in largescale studies, they are instrumental in optimizing the procedure and in devising efficient nearly exact pruning heuristics that can dramatically reduce the fraction of array entries that need to be computed (Havgaard et al., 2007).
The constructions presented here give rise to several variations. Point in case being the computation of hybrid probabilities, i.e., the probabilities $\mathbb{P}_{i, j ; h, \ell}^{\mathrm{Hy}}$ that $R[i, j]$ and $S[h, \ell]$ form an "interaction
stem" or a even an entire uninterrupted interaction region Huang et al. (2009). Another line of research concerns improved energy models for more complex types of loops Isambert and Siggia (2000).
The algorithmic approach taken here was motivated by a combinatorial analysis of zigzag-free interaction structures. From a mathematical point of view, our approach is centered around the notions of tight structures and their decomposition trees (the latter being described in the appendix). A detailed mathematical analysis, in particular the derivation of the generating function and further enumeration results, will be discussed elsewhere.
In order to store the partition function and the base pairing probabilities of joint structures in rip, we employ 4 -dimensional arrays. For the recursion for the partition function, $Q^{I}$, we use 16 matrices, 24 matrices for $Q^{R T}, 18$ matrices for $Q^{D T}$ and 45 matrices for $Q^{T}$, in the context of taking into account the loop energy. The complete set of partition function recursions and all details on the particular implementation of rip can be found at http://www.combinatorics.cn/cbpc/rip.html. The space complexity of rip is $O\left(N^{4}\right)$. Summations in our recursion equations run over at most two independent indices. Therefore, the time complexity in rip is $O\left(N^{6}\right)$. In order to obtain the pairing probabilities we trace back in the decomposition tree. Thus, we have the same space complexity and time complexity as for calculating the partition function.

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## REFERENCES

Akutsu, T. (2000) Dynamic programming algorithms for RNA secondary structure prediction with pseudoknots. Disc. Appl. Math., 104, 45-62.
Alkan, C., Karakoc, E., Nadeau, J., Sahinalp, S. and Zhang, K. (2006) RNA-RNA interaction prediction and antisense RNA target search. J. Comput. Biol., 13, 267282.

Andronescu, M., Zhang, Z. and Condon, A. (2005) Secondary structure prediction of interacting RNA molecules. J. Mol. Biol., 345, 1101-1112.
Argaman, L. and Altuvia, S. (2000) fhlA repression by OxyS RNA: kissing complex formation at two sites results in a stable antisense-target RNA complex. J. Mol. Biol., 300, 1101-1112.
Bachellerie, J., Cavaillé, J. and Hüttenhofer, A. (2002) The expanding snoRNA world. Biochimie, 84, 775-790.
Banerjee, D. and Slack, F. (2002) Control of developmental timing by small temporal RNAs: a paradigm for RNA-mediated regulation of gene expression. Bioessays, 24, 119-129.
Benne, R. (1992) RNA editing in trypanosomes. the use of guide RNAs. Mol. Biol.Rep., 16, 217-227.
Bernhart, S., Tafer, H., Mückstein, U., Flamm, C., Stadler, P. and Hofacker, I. (2006) Partition function and base pairing probabilities of RNA heterodimers. Algorithms Mol. Biol., 1, 3-3.
Busch, A., Richter, A. and Backofen, R. (2008) IntaRNA: efficient prediction of bacterial sRNA targets incorporating target site accessibility and seed regions. Bioinformatics, 24, 2849-2856.
Cary, R. and Stormo, G. (1995) Graph-theoretic approach to RNA modeling using comparative data. Proc. Int. Conf. Intell. Syst. Mol. Biol., 3, 75-80.

Chitsaz, H., Salari, R., Sahinalp, S. and Backofen, R. (2009) A partition function algorithm for interacting nucleic acid strands. Bioinformatics, 25, i365-i373.
Dimitrov, R. A. and Zuker, M. (2004) Prediction of hybridization and melting for double-stranded nucleic acids. Biophys. J., 87, 215-226.
Dirks, R. and Pierce, N. (2003) A partition function algorithm for nucleoic acid secondary structure inluding pseudoknots. J. Comput. Chem., 24, 1664-1677.
Gabow, H. (1973) Implementation of algorithms for maximum matching on nonbipartite graphs. Ph.D. thesis, Stanford University, Stanford (California). 248p.
Gago, S., De la Peña, M. and Flores, R. (2005) A kissing-loop interaction in a hammerhead viroid RNA critical for its in vitro folding and in vivo viability. RNA, 11, 1073-1083.
Geissmann, T. and Touati, D. (2004) Hfq, a new chaperoning role: binding to messenger RNA determines access for small RNA regulator. EMBO J., 23, 396-405.
Giegerich, R. and Meyer, C. (2002) Lecture Notes In Computer Science, volume 2422, chapter Algebraic Dynamic Programming, pp. 349-364. Springer-Verlag.
Hackermüller, J., Meisner, N., Auer, M., Jaritz, M. and Stadler, P. (2005) The effect of RNA secondary structures on RNA-ligand binding and the modifier RNA mechanism: a quantitative model. Gene, 345, 3-12.
Havgaard, J., Torarinsson, E. and Gorodkin, J. (2007) Fast pairwise structural RNA alignments by pruning of the dynamical programming matrix. PLoS Comput. Biol., 3, 1896-1908.
Hofacker, I., Fontana, W., Stadler, P., Bonhoeffer, L., Tacker, M. and Schuster, P. (1994) Fast folding and comparison of RNA secondary structures. Monatsh. Chem., 125, 167-188.
Huang, F., Qin, J., Reidys, C. and Stadler, P. (2009) Target prediction for RNA-RNA interaction.
Isambert, H. and Siggia, E. (2000) Modeling RNA folding paths with pseudoknots: application to hepatitis delta virus ribozyme. Proc. Natl. Acad. Sci. USA, 97, 65156520.

Kugel, J. and Goodrich, J. (2007) An RNA transcriptional regulator templates its own regulatory RNA. Nat. Struct. Mol. Biol., 3, 89-90.
Majdalani, N., Hernandez, D. and Gottesman, S. (2002) Regulation and mode of action of the second small RNA activator of RpoS translation, RprA. Mol. Microbiol., 46, 813-826.
Mathews, D., Sabina, J., Zuker, M. and Turner, D. (1999) Expanded sequence dependence of thermodynamic parameters improves prediction of RNA secondary structure. J. Mol. Biol., 288, 911-940.
McCaskill, J. (1990) The equilibrium partition function and base pair binding probabilities for RNA secondary structure. Biopolymers, 29, 1105-1119.
McManus, M. and Sharp, P. (2002) Gene silencing in mammals by small interfering RNAs. Nature Reviews, 3, 737-747.
Meisner, N., Hackermüller, J., Uhl, V., Aszódi, A., Jaritz, M. and Auer, M. (2004) mRNA openers and closers: modulating AU-rich element-controlled mRNA stability by a molecular switch in mRNA secondary structure. Chembiochem., 5, 1432-1447.
Mneimneh, S. (2007) On the approximation of optimal structures for RNARNA interaction. IEEE/ACM Trans. Comp. Biol. Bioinf. In press, doi.ieeecomputersociety.org/10.1109/TCBB.2007.70258.
Mückstein, U., Tafer, H., Bernhard, S., Hernandez-Rosales, M., Vogel, J., Stadler, P. and Hofacker, I. (2008) Translational control by RNA-RNA interaction: Improved computation of RNA-RNA binding thermodynamics. In Elloumi, M., Küng, J., Linial, M., Murphy, R. F., Schneider, K. and Toma, C. T. (eds.), BioInformatics Research and Development - BIRD 2008, volume 13 of Comm. Comp. Inf. Sci., pp. 114-127. Springer, Berlin.
Mückstein, U., Tafer, H., Hackermüller, J., Bernhard, S., Stadler, P. and Hofacker, I. (2006) Thermodynamics of RNA-RNA binding. Bioinformatics, 22, 1177-1182. Earlier version in: German Conference on Bioinformatics 2005, Torda andrew and Kurtz, Stefan and Rarey, Matthias (eds.), Lecture Notes in Informatics P-71, pp 3-13, Gesellschaft f. Informatik, Bonn 2005.
Narberhaus, F. and Vogel, J. (2007) Sensory and regulatory RNAs in prokaryotes: A new german research focus. RNA Biol., 4, 160-164.
Pervouchine, D. (2004) IRIS: Intermolecular RNA interaction search. Proc. Genome Informatics, 15, 92-101.
Qin, J. and Reidys, C. (2008) A framework for RNA tertiary interaction. Submitted.
Rehmsmeier, M., Steffen, P., Höchsmann, M. and Giegerich, R. (2004) Fast and effective prediction of microRNA/target duplexes. Gene, 10, 1507-1517.
Ren, J., Rastegari, B., Condon, A. and Hoos, H. (2005) Hotknots: heuristic prediction of microRNA secondary structures including pseudoknots. RNA, 11, 1494-1504.
Rivas, E. and Eddy, S. (1999) A dynamic programming algorithms for RNA structure prediction including pseudoknots. J. Mol. Biol., 285, 2053-2068.

Sharma, C., Darfeuille, F., Plantinga, T. and Vogel, J. (2007) A small RNA regulates multiple ABC transporter mRNAs by targeting C/A-rich elements inside and upstream of ribosome-binding sites. Genes \& Dev., 21, 2804-2817.
Tafer, H., Kehr, S., Hertel, J. and Stadler, P. (2009) RNAsnoop: Efficient target prediction for box H/ACA snoRNAs. Submitted.

Zuker, M. and Stiegler, P. (1981) Optimal computer folding of large rna sequences using thermodynamics and auxiliary information. Nucleic Acids Res., 9, 133-148.

